CHIMIA 50 (1996) Nr. 6 (Juni)

ical number of compounds which are to be made in the particular library. Because the final product of this library is a discrete array, biological screening of individual compounds of known structure can provide an SAR profile of the complete library. The development of READ/WRITE strategies which further improve the efficiency of this approach are currently underway and will be reported in due course.

Experimental Procedure

The tea bags were apportioned and scanned as described in text and Fig. 1. a) Each of the four FMOC-amino acids (4.59 mmol), DIC (4.59 mmol), and HOBt (3.44 mmol) were dissolved in THF (10 ml) individually and stirred for 20 min, filtered, and the filtrate added to 1.275 g of Wang resin (Advanced Chemtech, 0.9 mmol/g) in tea bags containing the transponders (16/flask). After 18 h of gentle stirring, the resin was washed with DMF, MeOH, and CH_2Cl_2 . The tea bags were immersed in 20% piperidine/DMF for 30 min and then rinsed with MeOH and CH₂Cl₂, then scanned and sorted to four new reaction vessels. b) To each reaction vessel was added 15 ml of MeOH/CH₂Cl₂(1:1), one of four aldehydes (15 mmol), p-hydroxyphenylacetic acid (15 mmol), and benzyl isocyanide (15 mmol). The resin was gently stirred for 12 h, then rinsed repeatedly with MeOH and CH2Cl2 and sorted to four new reaction vessels. c) To each of the reaction vessels was added the acid (14 mmol) followed by DCC (15 mmol) in 15 ml of pyridine. After 18 h, the resin was rinsed with DMF (50°), then MeOH and CH_2Cl_2 . d) The tea bags were scanned and sorted into individual wells of a polypropylene microtiter plate (96 well, 2 ml/ well). A soln. (1 ml) of 20% TFA/CH₂Cl₂ was added to each well and let stand for 20 min. The tea bags were removed and rinsed with CH₂Cl₂ and the solvents stripped i.v.

This research was supported by NIH grant GM-20080.

Received: May 10, 1996

- [1] M. Gallop, R. Barrett, W. Dower, S. Fodor, E. Gordon, J. Med. Chem. 1994, 37, 1233; E. Gordon, R. Barrett, W. Dower, S. Fodor, M. Gallop, ibid. 1994, 37, 1385; S. Blondelle, E. Perezpaya, C. Dooley, C. Pinilla, R. Houghten, Trac-Trends Anal. Chem. 1995, 14, 83; M. Lebel, V. Krchnak, N. Sepetov, B. Seligmann, P. Strop, S. Felder, K. Lam, Biopolymers 1995, 37, 177; K. Janda, Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 10779; W. Moos, G. Green, M. Pavia, Ann. Rep. Med. Chem. 1993, 28, 315.
- [2] H. Geysen, R. Meloen, S. Barteling, Proc. Natl. Acad. Sci. U.S.A. 1984, 81, 3998; A. Gray, R. Balerio, A. Dispasquale, J. Greig, N. Maeji, J. Pept. Sci. 1995, 1, 80; S. Dewitt, J. Kiely, C. Stnakovic, M. Schroeder, D. Cody, M. Pavia, Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 6909.
- C. Holmes, C. Adams, L. Kochersperger, [3] R. Mortensen, L. Aldwin, Biopolymers 1995, 37, 199; S. Fodor, J. Read, M. Pirrung, L. Stryer, A. Lu, D. Solas, Science 1991. 251. 767.
- [4] F. Sebestyen, G. Dibo, A. Kovacs, A. Furka, Bio Med. Chem. Lett. 1993, 3, 413; K. Lam, S. Salmon, E. Hersh, V. Hruby, W. Kazmeierski, R. Knapp, Nature 1992, 360, 768; A. Furka, F. Sebestyen, M. Asgedom, G. Dibo, Int. J. Pept. Protein Res. 1991, 37, 487; R. Houghten, C. Pinilla, S. Blondelle, J. Appel, C. Dooley, J. Cuervo, Nature (London) 1991, 354, 84; K. Lam, S. Salmon, E. Hersh, V. Hruby, W. Dazmierski, R. Knapp, ibid. 1991, 354, 82.
- [5] C. Dooley, N. Chung, B. Wilkes, P. Schiller, J. Bidlack, G. Pasternak, R. Houghten, Science 1994, 266, 2019.
- S. Brenner, R. Lerner, Proc. Natl. Acad. Sci. U.S.A. 1992, 89, 5381; J. Kerr, S. [6] Banville, R. Zuchermann, J. Am. Chem. Soc. 1993, 115, 2529; M. Ohlmeyer, R. Swanson, L. Dillard, J. Reader, G. Asouline, R. Kobayashi, M. Wigler, W. Still, Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 10922.
- [7] a) U.S. Pat. No. 5,252,962, Bio Medic Data

Systems, Inc., Maywood, N. J.; b) U.S. Pat. No. 5,351,052, Texas Instruments, Inc., Austin, TX.

- [8] Implantable Micro Identification (IMI) systems, Bio Medic Data Sytems, Inc. Maywood, N.J. Transponders were scanned using a DAS-4004 Pocket scanner and the ID's can be downloaded to a Mac or PC.
- [9] Transponders were scanned using a DAS-4004 Pocket scanner.
- [10] S.-S. Wang, J. Am. Chem. Soc. 1973, 95, 1328; G. Lu, S. Mojsov, J. Tam, R. Merrifield, J. Org. Chem. 1981, 46, 3433.
- [11] Tea bags can be combined such that compound mixtures of known composition can be screened.
- [12] Yields were obtained from the weight of individual products in library after removal from resin. They are based on initial loading of polymer as described by manufacturer and represent the overall yield for the five steps shown in the Scheme. Low-resolution electrospray MS was obtained for all compounds. Full characterization (HRMS. ¹³C- and ¹H-NMR, IR) of several members $(A_1B_4C_1, A_4B_1C_3)$ was obtained: $A_1B_1C_1$, 50%; A₁B₁C₂, 45%; A₁B₁C₃, 49%; A₁B₁C₄, $39\%; A_1B_2C_1, 55\%; A_1B_2C_2, 53\%; A_1B_2C_3,$ $64\%; A_1B_2C_4, 76\%; A_1B_3C_1, 49\%; A_1B_3C_2,\\$ 51%; A₁B₃C₃, 61%; A₁B₃C₄, 67%; A₁B₄C₁, $50\%; A_1B_4C_2, 50\%; A_1B_4C_3, 57\%; A_1B_4C_4,$ $40\%; A_2B_1C_1, 44\%, A_2B_1C_2, 44\%; A_2B_1C_3,$ $52\%; A_2B_1C_4, 56\%; A_2B_2C_1, 40\%, A_2B_2C_2,$ 43%, A₂B₂C₃, 52%; A₂B₂C₄, 55%; A₂B₃C₁, 43%, A₂B₃C₂, 36%; A₂B₃C₃, 42%; A₂B₃C₄, $48\%; A_2B_4C_1, 40\%; A_2B_4C_2, 37\%; A_2B_4C_3,$ 44%; A₂B₄C₄, 57%; A₃B₁C₁, 65%; A₃B₁C₂, $68\%; A_3B_1C_3, 73\%; A_3B_1C_4, 60\%; A_3B_2C_1,$ 30%; A₃B₂C₂, 64%; A₃B₂C₃, 69%; A₃B₂C₄, $76\%; A_3B_3C_1, 54\%; A_3B_3C_2, 55\%; A_3B_3C_3,$ 58%; A₃B₃C₄, 30%; A₃B₄C₁, 55%; A₃B₄C₂, 58%; A3B4C3, 64%; A3B4C4, 66%; A4B1C1, $61\%; A_4B_1C_2, 61\%; A_4B_1C_3, 71\%; A_4B_1C_4,$ $72\%; A_4B_2C_1, 55\%; A_4B_2C_2, 56\%; A_4B_2C_3,\\$ 57%; A₄B₂C₄, 61%; A₄B₃C₁, 47%; A₄B₃C₂, 46%; A4B3C3, 47%; A4B3C4, 54%; A4B4C1, 55%; A₄B₄C₂, 49%; A₄B₄C₃, 59%; A₄B₄C₄, 45%.

Chimia 50 (1996) 260-261 © Neue Schweizerische Chemische Gesellschaft ISSN 0009-4293

The Solid-Phase Synthesis of **Complex Small Molecules**

Jonathan A. Ellman*

of small-molecule libraries currently being performed in our laboratory is overviewed. We consider a number of factors in the selection of a compound class for library synthesis. One strategy that we have employed is to select 'privileged'

The design, synthesis, and evaluation structures, where the display of different functionality upon the structure has previously provided a number of potent and specific drugs or candidates towards different therapeutic targets. The first class of 'privileged' structures that we focused on were the 1,4-benzodiazepines, one of the most prescribed classes of orally active drugs that target a wide range of different receptors and enzymes. Other examples include libraries based upon tropane, prostaglandin, and tricyclic frameworks. An alternative strategy that we have employed is to design compound classes based on important biological recognition motifs. One example where we have applied this strategy is the synthesis of libraries of mimetics of β -turns, which play a key role in molecular recognition events in biological systems. A second example is the

*Correspondence: Prof. J.A. Ellman College of Chemistry University of California Berkeley Berkeley, CA 94720, USA

CHIMIA 50 (1996) Nr. 6 (Juni)

ical number of compounds which are to be made in the particular library. Because the final product of this library is a discrete array, biological screening of individual compounds of known structure can provide an SAR profile of the complete library. The development of READ/WRITE strategies which further improve the efficiency of this approach are currently underway and will be reported in due course.

Experimental Procedure

The tea bags were apportioned and scanned as described in text and Fig. 1. a) Each of the four FMOC-amino acids (4.59 mmol), DIC (4.59 mmol), and HOBt (3.44 mmol) were dissolved in THF (10 ml) individually and stirred for 20 min, filtered, and the filtrate added to 1.275 g of Wang resin (Advanced Chemtech, 0.9 mmol/g) in tea bags containing the transponders (16/flask). After 18 h of gentle stirring, the resin was washed with DMF, MeOH, and CH_2Cl_2 . The tea bags were immersed in 20% piperidine/DMF for 30 min and then rinsed with MeOH and CH₂Cl₂, then scanned and sorted to four new reaction vessels. b) To each reaction vessel was added 15 ml of MeOH/CH₂Cl₂(1:1), one of four aldehydes (15 mmol), p-hydroxyphenylacetic acid (15 mmol), and benzyl isocyanide (15 mmol). The resin was gently stirred for 12 h, then rinsed repeatedly with MeOH and CH2Cl2 and sorted to four new reaction vessels. c) To each of the reaction vessels was added the acid (14 mmol) followed by DCC (15 mmol) in 15 ml of pyridine. After 18 h, the resin was rinsed with DMF (50°), then MeOH and CH_2Cl_2 . d) The tea bags were scanned and sorted into individual wells of a polypropylene microtiter plate (96 well, 2 ml/ well). A soln. (1 ml) of 20% TFA/CH₂Cl₂ was added to each well and let stand for 20 min. The tea bags were removed and rinsed with CH₂Cl₂ and the solvents stripped i.v.

This research was supported by NIH grant GM-20080.

Received: May 10, 1996

- [1] M. Gallop, R. Barrett, W. Dower, S. Fodor, E. Gordon, J. Med. Chem. 1994, 37, 1233; E. Gordon, R. Barrett, W. Dower, S. Fodor, M. Gallop, ibid. 1994, 37, 1385; S. Blondelle, E. Perezpaya, C. Dooley, C. Pinilla, R. Houghten, Trac-Trends Anal. Chem. 1995, 14, 83; M. Lebel, V. Krchnak, N. Sepetov, B. Seligmann, P. Strop, S. Felder, K. Lam, Biopolymers 1995, 37, 177; K. Janda, Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 10779; W. Moos, G. Green, M. Pavia, Ann. Rep. Med. Chem. 1993, 28, 315.
- [2] H. Geysen, R. Meloen, S. Barteling, Proc. Natl. Acad. Sci. U.S.A. 1984, 81, 3998; A. Gray, R. Balerio, A. Dispasquale, J. Greig, N. Maeji, J. Pept. Sci. 1995, 1, 80; S. Dewitt, J. Kiely, C. Stnakovic, M. Schroeder, D. Cody, M. Pavia, Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 6909.
- C. Holmes, C. Adams, L. Kochersperger, [3] R. Mortensen, L. Aldwin, Biopolymers 1995, 37, 199; S. Fodor, J. Read, M. Pirrung, L. Stryer, A. Lu, D. Solas, Science 1991. 251. 767.
- [4] F. Sebestyen, G. Dibo, A. Kovacs, A. Furka, Bio Med. Chem. Lett. 1993, 3, 413; K. Lam, S. Salmon, E. Hersh, V. Hruby, W. Kazmeierski, R. Knapp, Nature 1992, 360, 768; A. Furka, F. Sebestyen, M. Asgedom, G. Dibo, Int. J. Pept. Protein Res. 1991, 37, 487; R. Houghten, C. Pinilla, S. Blondelle, J. Appel, C. Dooley, J. Cuervo, Nature (London) 1991, 354, 84; K. Lam, S. Salmon, E. Hersh, V. Hruby, W. Dazmierski, R. Knapp, ibid. 1991, 354, 82.
- [5] C. Dooley, N. Chung, B. Wilkes, P. Schiller, J. Bidlack, G. Pasternak, R. Houghten, Science 1994, 266, 2019.
- S. Brenner, R. Lerner, Proc. Natl. Acad. Sci. U.S.A. 1992, 89, 5381; J. Kerr, S. [6] Banville, R. Zuchermann, J. Am. Chem. Soc. 1993, 115, 2529; M. Ohlmeyer, R. Swanson, L. Dillard, J. Reader, G. Asouline, R. Kobayashi, M. Wigler, W. Still, Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 10922.
- [7] a) U.S. Pat. No. 5,252,962, Bio Medic Data

Systems, Inc., Maywood, N. J.; b) U.S. Pat. No. 5,351,052, Texas Instruments, Inc., Austin, TX.

- [8] Implantable Micro Identification (IMI) systems, Bio Medic Data Sytems, Inc. Maywood, N.J. Transponders were scanned using a DAS-4004 Pocket scanner and the ID's can be downloaded to a Mac or PC.
- [9] Transponders were scanned using a DAS-4004 Pocket scanner.
- [10] S.-S. Wang, J. Am. Chem. Soc. 1973, 95, 1328; G. Lu, S. Mojsov, J. Tam, R. Merrifield, J. Org. Chem. 1981, 46, 3433.
- [11] Tea bags can be combined such that compound mixtures of known composition can be screened.
- [12] Yields were obtained from the weight of individual products in library after removal from resin. They are based on initial loading of polymer as described by manufacturer and represent the overall yield for the five steps shown in the Scheme. Low-resolution electrospray MS was obtained for all compounds. Full characterization (HRMS. ¹³C- and ¹H-NMR, IR) of several members $(A_1B_4C_1, A_4B_1C_3)$ was obtained: $A_1B_1C_1$, 50%; A₁B₁C₂, 45%; A₁B₁C₃, 49%; A₁B₁C₄, $39\%; A_1B_2C_1, 55\%; A_1B_2C_2, 53\%; A_1B_2C_3,$ $64\%; A_1B_2C_4, 76\%; A_1B_3C_1, 49\%; A_1B_3C_2,\\$ 51%; A₁B₃C₃, 61%; A₁B₃C₄, 67%; A₁B₄C₁, $50\%; A_1B_4C_2, 50\%; A_1B_4C_3, 57\%; A_1B_4C_4,$ $40\%; A_2B_1C_1, 44\%, A_2B_1C_2, 44\%; A_2B_1C_3,$ $52\%; A_2B_1C_4, 56\%; A_2B_2C_1, 40\%, A_2B_2C_2,$ 43%, A₂B₂C₃, 52%; A₂B₂C₄, 55%; A₂B₃C₁, 43%, A₂B₃C₂, 36%; A₂B₃C₃, 42%; A₂B₃C₄, 48%; A₂B₄C₁, 40%; A₂B₄C₂, 37%; A₂B₄C₃, 44%; A₂B₄C₄, 57%; A₃B₁C₁, 65%; A₃B₁C₂, $68\%; A_3B_1C_3, 73\%; A_3B_1C_4, 60\%; A_3B_2C_1,$ 30%; A₃B₂C₂, 64%; A₃B₂C₃, 69%; A₃B₂C₄, $76\%; A_3B_3C_1, 54\%; A_3B_3C_2, 55\%; A_3B_3C_3,$ 58%; A₃B₃C₄, 30%; A₃B₄C₁, 55%; A₃B₄C₂, 58%; A3B4C3, 64%; A3B4C4, 66%; A4B1C1, $61\%; A_4B_1C_2, 61\%; A_4B_1C_3, 71\%; A_4B_1C_4,$ $72\%; A_4B_2C_1, 55\%; A_4B_2C_2, 56\%; A_4B_2C_3,\\$ 57%; A₄B₂C₄, 61%; A₄B₃C₁, 47%; A₄B₃C₂, 46%; A4B3C3, 47%; A4B3C4, 54%; A4B4C1, 55%; A₄B₄C₂, 49%; A₄B₄C₃, 59%; A₄B₄C₄, 45%.

Chimia 50 (1996) 260-261 © Neue Schweizerische Chemische Gesellschaft ISSN 0009-4293

The Solid-Phase Synthesis of **Complex Small Molecules**

Jonathan A. Ellman*

of small-molecule libraries currently being performed in our laboratory is overviewed. We consider a number of factors in the selection of a compound class for library synthesis. One strategy that we have employed is to select 'privileged'

The design, synthesis, and evaluation structures, where the display of different functionality upon the structure has previously provided a number of potent and specific drugs or candidates towards different therapeutic targets. The first class of 'privileged' structures that we focused on were the 1,4-benzodiazepines, one of the most prescribed classes of orally active drugs that target a wide range of different receptors and enzymes. Other examples include libraries based upon tropane, prostaglandin, and tricyclic frameworks. An alternative strategy that we have employed is to design compound classes based on important biological recognition motifs. One example where we have applied this strategy is the synthesis of libraries of mimetics of β -turns, which play a key role in molecular recognition events in biological systems. A second example is the

*Correspondence: Prof. J.A. Ellman College of Chemistry University of California Berkeley Berkeley, CA 94720, USA

261

synthesis of libraries of compounds that are based on an isostere that mimics the tetrahedral intermediate for peptide hydrolysis as catalyzed by the therapeutically important aspartic-acid class of proteases.

For each library synthesis project the synthesis approach is designed to achieve three goals: 1) Several different buildingblock sets should be incorporated to provide rapid access to a large number of diverse compounds. 2) The chemistry should be compatible with the display of as much functionality as possible including reactive functionality that is commonly found in drugs such as alcohols, phenols, indoles, carboxylic acids, amides, nitriles, nitro groups, and halides. 3) The building blocks used in the synthesis of the library should be commercially available or at least readily accessible to facilitate rapid library synthesis. A considerable number of synthesis methods are explored in order to achieve these goals. Two main areas of focus are C–C-bond forming processes and linkage strategies for attaching compounds to the solid support. Some of the solid-phase chemistry that is developed is based upon analogous chemistry in solution; however, much of chemistry that is developed does not have a current solution-phase counterpart.

According to the above strategies and goals we have generated libraries of thousands to tens of thousands of compounds. Library synthesis is performed in a microtiter plate format either using the *Chiron Mimotope* pin method initially developed by *Geysen* for peptide epitope mapping or by analogous bead-based approaches. Evaluation of these compound libraries has lead to the identification of numerous ligands to different receptor targets to serve as tools to probe receptor function. Selected examples include the identification

Chimia 50 (1996) 261–266 © Neue Schweizerische Chemische Gesellschaft ISSN 0009–4293

The Solid-Phase Part of Supported Small-Molecule Synthesis

Mark J. Kurth*

Abstract. The synthesis of small molecules on solid phase must not only address the vagaries of C–C-bond formation and functional-group manipulation, but must also take into account solid-support issues such as 'point of attachment', 'resin compatibility', 'reagent accesibility', and 'product liberation'. Hence, the resin plays a vital role in the solid-phase venture and the polymer advantages can be summarized as reactions can be driven to completion by addition of excess solution-phase reagents, reaction products are 'isolated' by filtration and washing, and multiple-step synthesis terminating with a 'selective' liberation step can deliver essentially pure product. These issues, as well as a number of strategies for the preparation and functionalization of resin supports, are discussed.

Introduction

Adapting solution-phase organic reactions to solid-phase techniques is one of the important challenges embraced by the burgeoning field of small-molecule combinatorial chemistry. In a solid-phase arena, strategic synthetic planning must not only address the vagaries of C–C-bond formation and functional-group manipulation, but must also take into account solid-support issues such as 'point of attachment', 'resin compatibility', 'reagent accessibility', and 'product liberation'.

The Polymer Advantage

The pioneering small-molecule solidphase work of *Leznoff* [1] and the more recent efforts of many academic and industrial chemists [2] have established three principal synthetic advantages of solidphase techniques (*i.e.*, the *polymer advantage*): *i*) many solid-phase reactions can be driven to completion by addition of excess solution-phase reagents (removed by filtration and washing), *ii*) solid-phase reaction products are 'isolated' by filtration and washing, and *iii*) multiple-step of benzodiazepine-based inhibitors of pp60^{src} tyrosine kinase, which is implicated in cancer, in collaboration with *Victor Levine* at the *M.D. Anderson* Cancer Institute, benzodiazepine-based ligands to DNA-binding autoantibodies implicated in the autoimmune disease systemic *lupus erythematosus* which are currently being evaluated in mouse models of the disease in collaboration with *Gary Glick* at the University of Michigan, and hydroxyethylamine isostere-based nonpeptide inhibitors of cathepsin D with low nanomolar Ki values.

Received: May 15, 1996

[1] J.A. Ellman, Acc. Chem. Res. 1996, 29, 132.

- [2] N.J. Maeji, R.M. Valerio, A.M. Bray, R.A. Campbell, H.M. Geysen, *Reactive Polym.* 1994, 22, 203.
- [3] H.M. Geysen, S.J. Rodda, T.J. Mason, G. Tribbick, P.G. Schoofs, J. Immunol. Methods 1987, 102, 259.

synthesis terminating with a 'selective' liberation step can deliver essentially pure product.

Collectively, these advantages suggest that the solid-support can play a much broader role in solid-phase chemistry than merely that of an inert matrix. When appropriately designed, its implications can be far reaching and can elevate solidphase techniques to exceptional synthetic advantage. Consider the sequent-auxiliary concept [3] outlined in Scheme 1 where a polymer-bound chiral auxiliary (AUX) is called upon to mediate a diastereoselective C α -alkylation reaction in Step 1 and a diastereoselective iodolactonization reaction in Step 2. Thus, in addition to embracing the polymer advantage, there is significantly improved synthetic economy and atom efficiency in that the chiral auxiliary is called upon to mediate two diastereoselective transformation, and the chiral auxiliary can be recovered by simple filtration.

To effect this strategy, C_2 -symmetric pyrrolidine-based auxiliary 1 was attached to *Merrifield* resin (\mathbb{B} = polystyrene) and the remaining OH group blocked by benzylation to give the pseudo- C_2 -symmetric auxiliary **2**. AUX-Mediated C α -alkylation (93.5:6.5 diastereoselectivity) followed by AUX-mediated iodolactonization (>99:1 diastereoselectivity) delivered targeted lactone **4** (R = CH₃) essentially

*Correspondence: Prof. M.J. Kurth Department of Chemistry University of California, Davis Davis, California 95616, USA

261

synthesis of libraries of compounds that are based on an isostere that mimics the tetrahedral intermediate for peptide hydrolysis as catalyzed by the therapeutically important aspartic-acid class of proteases.

For each library synthesis project the synthesis approach is designed to achieve three goals: 1) Several different buildingblock sets should be incorporated to provide rapid access to a large number of diverse compounds. 2) The chemistry should be compatible with the display of as much functionality as possible including reactive functionality that is commonly found in drugs such as alcohols, phenols, indoles, carboxylic acids, amides, nitriles, nitro groups, and halides. 3) The building blocks used in the synthesis of the library should be commercially available or at least readily accessible to facilitate rapid library synthesis. A considerable number of synthesis methods are explored in order to achieve these goals. Two main areas of focus are C–C-bond forming processes and linkage strategies for attaching compounds to the solid support. Some of the solid-phase chemistry that is developed is based upon analogous chemistry in solution; however, much of chemistry that is developed does not have a current solution-phase counterpart.

According to the above strategies and goals we have generated libraries of thousands to tens of thousands of compounds. Library synthesis is performed in a microtiter plate format either using the *Chiron Mimotope* pin method initially developed by *Geysen* for peptide epitope mapping or by analogous bead-based approaches. Evaluation of these compound libraries has lead to the identification of numerous ligands to different receptor targets to serve as tools to probe receptor function. Selected examples include the identification

Chimia 50 (1996) 261–266 © Neue Schweizerische Chemische Gesellschaft ISSN 0009–4293

The Solid-Phase Part of Supported Small-Molecule Synthesis

Mark J. Kurth*

Abstract. The synthesis of small molecules on solid phase must not only address the vagaries of C–C-bond formation and functional-group manipulation, but must also take into account solid-support issues such as 'point of attachment', 'resin compatibility', 'reagent accesibility', and 'product liberation'. Hence, the resin plays a vital role in the solid-phase venture and the polymer advantages can be summarized as reactions can be driven to completion by addition of excess solution-phase reagents, reaction products are 'isolated' by filtration and washing, and multiple-step synthesis terminating with a 'selective' liberation step can deliver essentially pure product. These issues, as well as a number of strategies for the preparation and functionalization of resin supports, are discussed.

Introduction

Adapting solution-phase organic reactions to solid-phase techniques is one of the important challenges embraced by the burgeoning field of small-molecule combinatorial chemistry. In a solid-phase arena, strategic synthetic planning must not only address the vagaries of C–C-bond formation and functional-group manipulation, but must also take into account solid-support issues such as 'point of attachment', 'resin compatibility', 'reagent accessibility', and 'product liberation'.

The Polymer Advantage

The pioneering small-molecule solidphase work of *Leznoff* [1] and the more recent efforts of many academic and industrial chemists [2] have established three principal synthetic advantages of solidphase techniques (*i.e.*, the *polymer advantage*): *i*) many solid-phase reactions can be driven to completion by addition of excess solution-phase reagents (removed by filtration and washing), *ii*) solid-phase reaction products are 'isolated' by filtration and washing, and *iii*) multiple-step of benzodiazepine-based inhibitors of pp60^{src} tyrosine kinase, which is implicated in cancer, in collaboration with *Victor Levine* at the *M.D. Anderson* Cancer Institute, benzodiazepine-based ligands to DNA-binding autoantibodies implicated in the autoimmune disease systemic *lupus erythematosus* which are currently being evaluated in mouse models of the disease in collaboration with *Gary Glick* at the University of Michigan, and hydroxyethylamine isostere-based nonpeptide inhibitors of cathepsin D with low nanomolar Ki values.

Received: May 15, 1996

[1] J.A. Ellman, Acc. Chem. Res. 1996, 29, 132.

- [2] N.J. Maeji, R.M. Valerio, A.M. Bray, R.A. Campbell, H.M. Geysen, *Reactive Polym.* 1994, 22, 203.
- [3] H.M. Geysen, S.J. Rodda, T.J. Mason, G. Tribbick, P.G. Schoofs, J. Immunol. Methods 1987, 102, 259.

synthesis terminating with a 'selective' liberation step can deliver essentially pure product.

Collectively, these advantages suggest that the solid-support can play a much broader role in solid-phase chemistry than merely that of an inert matrix. When appropriately designed, its implications can be far reaching and can elevate solidphase techniques to exceptional synthetic advantage. Consider the sequent-auxiliary concept [3] outlined in Scheme 1 where a polymer-bound chiral auxiliary (AUX) is called upon to mediate a diastereoselective C α -alkylation reaction in Step 1 and a diastereoselective iodolactonization reaction in Step 2. Thus, in addition to embracing the polymer advantage, there is significantly improved synthetic economy and atom efficiency in that the chiral auxiliary is called upon to mediate two diastereoselective transformation, and the chiral auxiliary can be recovered by simple filtration.

To effect this strategy, C_2 -symmetric pyrrolidine-based auxiliary 1 was attached to *Merrifield* resin (\mathbb{B} = polystyrene) and the remaining OH group blocked by benzylation to give the pseudo- C_2 -symmetric auxiliary **2**. AUX-Mediated C α -alkylation (93.5:6.5 diastereoselectivity) followed by AUX-mediated iodolactonization (>99:1 diastereoselectivity) delivered targeted lactone **4** (R = CH₃) essentially

*Correspondence: Prof. M.J. Kurth Department of Chemistry University of California, Davis Davis, California 95616, USA