

icry and artificial proteins. The state-of-the-art in chemoselective ligation and orthogonal protecting techniques determines the number of selectively attachable loops and thus the complexity of the molecules including TASP libraries for functional screening. However, the exploitation of the immense repertoire of synthetic organic chemistry for the incorporation of functional groups ('sticky ends') into peptide chains will rapidly expand the scope of the present approach. For example, ligand-directed assembly of helices, β -sheets, and loops onto tailor-made templates represents a powerful tool for studying supramolecular assembly and molecular recognition processes. Furthermore, the concepts described above open the way for a new generation of functional protein mimetics with a pivotal role in drug design.

This work was supported by the Swiss National Science Foundation.

Received: September 30, 1996

- [1] M. Mutter, *Angew. Chem. Int. Ed.* **1985**, *24*, 639.
- [2] J.S. Richardson, D.C. Richardson, *Trends Biochem. Sci.* **1989**, *14*, 304.
- [3] J.W. Bryson, S.F. Betz, H.S. Lu, D.J. Suich, H.X. Zhou, K.T. O'Neil, W.F. DeGrado, *Science* **1995**, *270*, 935.
- [4] J.S. Richardson, *Adv. Prot. Chem.* **1981**, *34*, 167.
- [5] S.R. Presnell, F.E. Cohen, *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 6592.
- [6] H. Li, R. Helling, C. Tang, N. Wingreen, *Science* **1966**, *273*, 666.
- [7] S. Borman, *Chem. Eng. News*, May **1996**, *27*, 29.
- [8] M. Mutter, G. Tuchscherer, *Makromol. Chem. Rapid Commun.* **1988**, *9* (6), 437.
- [9] M. Mutter, R. Hersperger, K. Gubernator, K. Mueller, *Proteins* **1989**, *5*, 13.
- [10] M. Mutter, E. Altmann, K.-H. Altmann, R. Hersperger, P. Koziej, K. Nebel, G. Tuchscherer, *Helv. Chim. Acta* **1989**, *71*, 835.
- [11] M. Mutter, S. Vuilleumier, *Angew. Chem. Int. Ed.* **1989**, *28*, 535.
- [12] G. Tuchscherer, M. Mutter, *J. Peptide Sci.* **1995**, *1*, 3.
- [13] G. Tuchscherer, M. Mutter, *J. Biotechnol.* **1995**, *41*, 197.
- [14] M. Mutter, G. Tuchscherer, C. Miller, K.-H. Altmann, R.J. Carey, D. Wyss, A.M. Labhardt, J. Rivier, *J. Am. Chem. Soc.* **1992**, *114*, 1463.
- [15] S. Anderson, H.L. Anderson, J.K.M. Sanders, *Acc. Chem. Res.* **1993**, *26*, 469.
- [16] U. Sila, M. Mutter, *J. Mol. Recognition* **1995**, *8*, 29.
- [17] M. Mutter, *Trends Biochem. Sci.* **1989**, *13*, 260.
- [18] R. Flögel, M. Mutter, *Biopolymers* **1992**, *32*, 1283.
- [19] I. Ernest, J. Kalvoda, C. Sigel, G. Rihs, H. Fritz, M.J.J. Blommers, F. Raschdorf, E. Francotte, M. Mutter, *Helv. Chim. Acta* **1993**, *76*, 1539.
- [20] P. Dumy, I.M. Eggleston, S.E. Cervigni, U. Sila, X. Sun, M. Mutter, *Tetrahedron Lett.* **1995**, *36*, 1255.
- [21] P. Dumy, I.M. Eggleston, G. Esposito, S. Nicula, M. Mutter, *Biopolymers*, in press.
- [22] I.M. Eggleston, M. Mutter, *Macromol. Symp.* **1996**, *101*, 397.
- [23] J.M. Stewart, J.D. Young, 'Solid Phase Peptide Synthesis', Pierce Chemical, 1984.
- [24] P. Lloyd-Williams, F. Albericio, F. Giralt, *Int. J. Pept. Protein Res.* **1991**, *37*, 58.
- [25] L.A. Vilaseca, K. Rose, R. Werlen, A. Meunir, R.E. Offord, C.L. Nichols, W.L. Scott, *Bioconj. Chem.* **1993**, *4*, 515.
- [26] C.-F. Liu, J.P. Tam, *J. Am. Chem. Soc.* **1994**, *116*, 4149.
- [27] P.E. Dawson, S.B.H. Kent, *J. Am. Chem. Soc.* **1993**, *115*, 7263.
- [28] G. Tuchscherer, *Tetrahedron Lett.* **1993**, *34*, 8419.
- [29] T. Haack, M. Mutter, *Tetrahedron Lett.* **1992**, *33*, 1589.
- [30] A. Nefzi, K. Schenk, M. Mutter, *Protein Peptide Lett.* **1994**, *1*, 66.
- [31] M. Mutter, A. Nefzi, T. Sato, X. Sun, F. Wahl, T. Wöhr, *Pept. Res.* **1995**, *8*, 145.
- [32] T. Wöhr, A. Nefzi, B. Rohwedder, T. Sato, X. Sun, F. Wahl, M. Mutter, *J. Am. Chem. Soc.*, in press.
- [33] P. Dumy, M. Keller, D.E. Ryan, B. Rohwedder, T. Wöhr, M. Mutter, *J. Am. Chem. Soc.*, in press.
- [34] A. Nefzi, X. Sun, M. Mutter, *Tetrahedron Lett.* **1995**, *36*, 229.
- [35] K.S. Ackerfeldt, R.M. Kim, D. Camac, J.D. Groves, J.D. Lear, W.F. DeGrado, *J. Am. Chem. Soc.* **1992**, *114*, 9656.
- [36] A. Grove, M. Mutter, J.E. Rivier, M. Montal, *J. Am. Chem. Soc.* **1993**, *115*, 1872.
- [37] M. Pawlak, U. Meseth, B. Dhanapal, M. Mutter, H. Vogel, *Protein Sci.* **1994**, *3*, 1788.
- [38] H. Vogel, M. Mutter *et al.*, in preparation.
- [39] E. Grouzmann, D. Felix, H. Imboden, A. Razaname, M. Mutter, *Eur. J. Biochem.* **1995**, *234*, 44.
- [40] E. Grouzmann, T. Buclin, M. Martire, B. Dörner, A. Razaname, M. Mutter, *Eur. J. Biochem.*, in press.
- [41] G. Tuchscherer, B. Dörner, U. Sila, B. Kamber, M. Mutter, *Tetrahedron* **1993**, *49*, 3559.
- [42] M. Mutter, P. Dumy, P. Garrouste, C. Lehmann, M. Mathieu, C. Peggion, S. Peluso, A. Razaname, G. Tuchscherer, *Angew. Chem. Int. Ed.* **E1996**, *35*, 1482.
- [43] P.G. Gerber, K. Müller, *J. Comput. Aided Mol. Des.* **1995**, *9*, 251.

Chimia 50 (1996) 648–649
© Neue Schweizerische Chemische Gesellschaft
ISSN 0009–4293

Azasugar Glycosidase Inhibitors: Useful Tools for Glycobiologists

Sylviane Picasso*

Abstract. Glycosidase inhibitors are moving increasingly as potential agents for the treatment of diseases by altering glycosylation or catabolism of glycoconjugates. The azasugars, a potent class of inhibitors have received much attention as tools for therapeutic investigation and in understanding of certain biological processes such as glycoprotein processing and enzymatic mechanisms. In this paper, we present some new synthetic azasugar compounds which show interesting activity as glycosidase inhibitors.

Introduction

Glycoproteins have many important biological functions: receptors, carriers, structural proteins, enzymes, hormones, lectins, and antibodies. Inhibitors of glycosidases, key enzymes in glycoprotein processing, are thus important as potential antiviral, antiadhesive, antitumoral, antibacterial, antihyperglycemic, or immunostimulatory agents [1]. Thus, an intense interest in the chemistry, biochemistry and pharmacology of glycosidase inhibitors has grown during the last decade and has

*Correspondence: Dr. S. Picasso
Institut de chimie organique
Université de Lausanne
CH-1015 Lausanne

icry and artificial proteins. The state-of-the-art in chemoselective ligation and orthogonal protecting techniques determines the number of selectively attachable loops and thus the complexity of the molecules including TASP libraries for functional screening. However, the exploitation of the immense repertoire of synthetic organic chemistry for the incorporation of functional groups ('sticky ends') into peptide chains will rapidly expand the scope of the present approach. For example, ligand-directed assembly of helices, β -sheets, and loops onto tailor-made templates represents a powerful tool for studying supramolecular assembly and molecular recognition processes. Furthermore, the concepts described above open the way for a new generation of functional protein mimetics with a pivotal role in drug design.

This work was supported by the Swiss National Science Foundation.

Received: September 30, 1996

- [1] M. Mutter, *Angew. Chem. Int. Ed.* **1985**, *24*, 639.
- [2] J.S. Richardson, D.C. Richardson, *Trends Biochem. Sci.* **1989**, *14*, 304.
- [3] J.W. Bryson, S.F. Betz, H.S. Lu, D.J. Suich, H.X. Zhou, K.T. O'Neil, W.F. DeGrado, *Science* **1995**, *270*, 935.
- [4] J.S. Richardson, *Adv. Prot. Chem.* **1981**, *34*, 167.
- [5] S.R. Presnell, F.E. Cohen, *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 6592.
- [6] H. Li, R. Helling, C. Tang, N. Wingreen, *Science* **1966**, *273*, 666.
- [7] S. Borman, *Chem. Eng. News*, May **1996**, *27*, 29.
- [8] M. Mutter, G. Tuchscherer, *Makromol. Chem. Rapid Commun.* **1988**, *9* (6), 437.
- [9] M. Mutter, R. Hersperger, K. Gubernator, K. Mueller, *Proteins* **1989**, *5*, 13.
- [10] M. Mutter, E. Altmann, K.-H. Altmann, R. Hersperger, P. Koziej, K. Nebel, G. Tuchscherer, *Helv. Chim. Acta* **1989**, *71*, 835.
- [11] M. Mutter, S. Vuilleumier, *Angew. Chem. Int. Ed.* **1989**, *28*, 535.
- [12] G. Tuchscherer, M. Mutter, *J. Peptide Sci.* **1995**, *1*, 3.
- [13] G. Tuchscherer, M. Mutter, *J. Biotechnol.* **1995**, *41*, 197.
- [14] M. Mutter, G. Tuchscherer, C. Miller, K.-H. Altmann, R.J. Carey, D. Wyss, A.M. Labhardt, J. Rivier, *J. Am. Chem. Soc.* **1992**, *114*, 1463.
- [15] S. Anderson, H.L. Anderson, J.K.M. Sanders, *Acc. Chem. Res.* **1993**, *26*, 469.
- [16] U. Sila, M. Mutter, *J. Mol. Recognition* **1995**, *8*, 29.
- [17] M. Mutter, *Trends Biochem. Sci.* **1989**, *13*, 260.
- [18] R. Flögel, M. Mutter, *Biopolymers* **1992**, *32*, 1283.
- [19] I. Ernest, J. Kalvoda, C. Sigel, G. Rihs, H. Fritz, M.J.J. Blommers, F. Raschdorf, E. Francotte, M. Mutter, *Helv. Chim. Acta* **1993**, *76*, 1539.
- [20] P. Dumy, I.M. Eggleston, S.E. Cervigni, U. Sila, X. Sun, M. Mutter, *Tetrahedron Lett.* **1995**, *36*, 1255.
- [21] P. Dumy, I.M. Eggleston, G. Esposito, S. Nicula, M. Mutter, *Biopolymers*, in press.
- [22] I.M. Eggleston, M. Mutter, *Macromol. Symp.* **1996**, *101*, 397.
- [23] J.M. Stewart, J.D. Young, 'Solid Phase Peptide Synthesis', Pierce Chemical, 1984.
- [24] P. Lloyd-Williams, F. Albericio, F. Giralt, *Int. J. Pept. Protein Res.* **1991**, *37*, 58.
- [25] L.A. Vilaseca, K. Rose, R. Werlen, A. Meunir, R.E. Offord, C.L. Nichols, W.L. Scott, *Bioconj. Chem.* **1993**, *4*, 515.
- [26] C.-F. Liu, J.P. Tam, *J. Am. Chem. Soc.* **1994**, *116*, 4149.
- [27] P.E. Dawson, S.B.H. Kent, *J. Am. Chem. Soc.* **1993**, *115*, 7263.
- [28] G. Tuchscherer, *Tetrahedron Lett.* **1993**, *34*, 8419.
- [29] T. Haack, M. Mutter, *Tetrahedron Lett.* **1992**, *33*, 1589.
- [30] A. Nefzi, K. Schenk, M. Mutter, *Protein Peptide Lett.* **1994**, *1*, 66.
- [31] M. Mutter, A. Nefzi, T. Sato, X. Sun, F. Wahl, T. Wöhr, *Pept. Res.* **1995**, *8*, 145.
- [32] T. Wöhr, A. Nefzi, B. Rohwedder, T. Sato, X. Sun, F. Wahl, M. Mutter, *J. Am. Chem. Soc.*, in press.
- [33] P. Dumy, M. Keller, D.E. Ryan, B. Rohwedder, T. Wöhr, M. Mutter, *J. Am. Chem. Soc.*, in press.
- [34] A. Nefzi, X. Sun, M. Mutter, *Tetrahedron Lett.* **1995**, *36*, 229.
- [35] K.S. Ackerfeldt, R.M. Kim, D. Camac, J.D. Groves, J.D. Lear, W.F. DeGrado, *J. Am. Chem. Soc.* **1992**, *114*, 9656.
- [36] A. Grove, M. Mutter, J.E. Rivier, M. Montal, *J. Am. Chem. Soc.* **1993**, *115*, 1872.
- [37] M. Pawlak, U. Meseth, B. Dhanapal, M. Mutter, H. Vogel, *Protein Sci.* **1994**, *3*, 1788.
- [38] H. Vogel, M. Mutter *et al.*, in preparation.
- [39] E. Grouzmann, D. Felix, H. Imboden, A. Razaname, M. Mutter, *Eur. J. Biochem.* **1995**, *234*, 44.
- [40] E. Grouzmann, T. Buclin, M. Martire, B. Dörner, A. Razaname, M. Mutter, *Eur. J. Biochem.*, in press.
- [41] G. Tuchscherer, B. Dörner, U. Sila, B. Kamber, M. Mutter, *Tetrahedron* **1993**, *49*, 3559.
- [42] M. Mutter, P. Dumy, P. Garrouste, C. Lehmann, M. Mathieu, C. Peggion, S. Peluso, A. Razaname, G. Tuchscherer, *Angew. Chem. Int. Ed.* **E1996**, *35*, 1482.
- [43] P.G. Gerber, K. Müller, *J. Comput. Aided Mol. Des.* **1995**, *9*, 251.

Chimia 50 (1996) 648–649
© Neue Schweizerische Chemische Gesellschaft
ISSN 0009–4293

Azasugar Glycosidase Inhibitors: Useful Tools for Glycobiologists

Sylviane Picasso*

Abstract. Glycosidase inhibitors are moving increasingly as potential agents for the treatment of diseases by altering glycosylation or catabolism of glycoconjugates. The azasugars, a potent class of inhibitors have received much attention as tools for therapeutic investigation and in understanding of certain biological processes such as glycoprotein processing and enzymatic mechanisms. In this paper, we present some new synthetic azasugar compounds which show interesting activity as glycosidase inhibitors.

Introduction

Glycoproteins have many important biological functions: receptors, carriers, structural proteins, enzymes, hormones, lectins, and antibodies. Inhibitors of glycosidases, key enzymes in glycoprotein processing, are thus important as potential antiviral, antiadhesive, antitumoral, antibacterial, antihyperglycemic, or immunostimulatory agents [1]. Thus, an intense interest in the chemistry, biochemistry and pharmacology of glycosidase inhibitors has grown during the last decade and has

*Correspondence: Dr. S. Picasso
Institut de chimie organique
Université de Lausanne
CH-1015 Lausanne

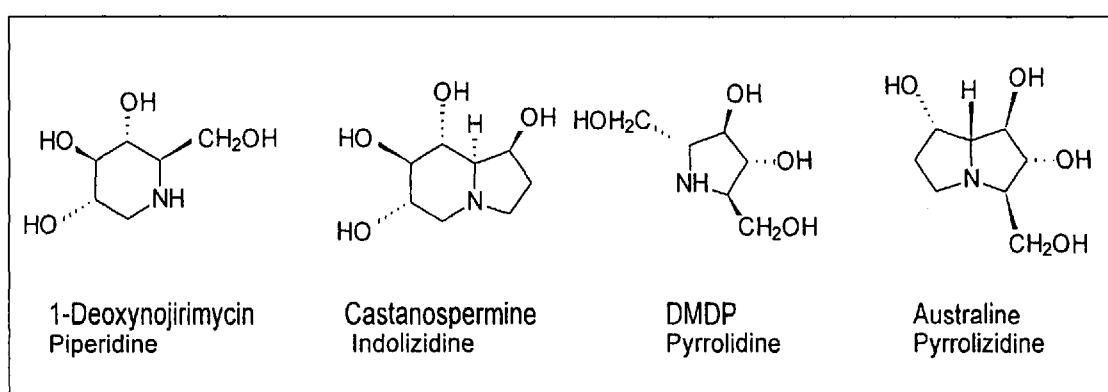


Fig. 1. Some azasugar glycosidase inhibitors

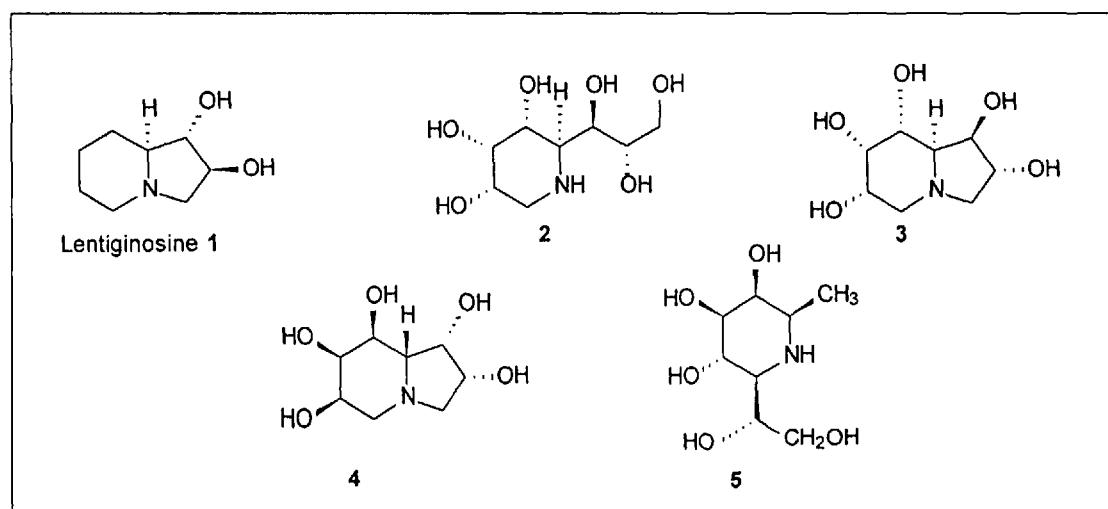


Fig. 2. New azasugar glycosidase inhibitors

led to a tremendous interest and demand for new compounds.

An important family of inhibitors are the azasugar compounds, analogues of monosaccharides in which the ring O-atom has been replaced by a N-atom. This renders the compound metabolically inert, but does not prevent its recognition by glycosidases due to mimicking of the furanose and pyranose moieties of the corresponding substrate. Azasugars include polyhydroxylated derivatives of piperidines, pyrrolidines, indolizidines, and pyrrolizidines (Fig. 1). Many have been isolated from plants and microorganisms and a number of synthetic analogues have been synthesized.

Activity of Some New Azasugars

Enzyme inhibition tests were performed on 24 commercial glycosidases with corresponding *p*-nitrophenyl glycoside substrates. IC_{50} values (concentration of inhibitor required for 50% inhibition of enzyme activity) and inhibition constants (K_i) were determined for some new azasugar compounds (Fig. 2).

Lentiginosine (1), a dihydroxylated indolizidine isolated from a natural source, is a powerful and specific inhibitor of

amyloglucosidases (1,4- α -D-glucan glycohydrolases, EC 3.2.1.3) [2]. The absolute configuration of the alkaloid was determined on the basis of amyloglucosidases inhibition tests with both lentiginosine enantiomers synthesized in Brandi's laboratory (University of Firenze). The enzymatic tests showed that the active enantiomer was the (+)-compound with a $K_i = 2 \mu\text{M}$. The absolute configuration of natural lentiginosine ((+)-1) was determined to be (1S,2S,8aS)-1,2-dihydroxyindolizidine [3]. Some compounds synthesized in our laboratory also displayed interesting glycosidase inhibition activity. The first 1,5-dideoxy-1,5-iminoctitols and the corresponding pentahydroxyindolizidines were prepared by Chen and Vogel [4]. The imino-octitol (2) and the pentahydroxyindolizidine (3) are better inhibitors of β -galactosidases than swainsonine (Fig. 1). The polyhydroxyindolizidine (4) is, like swainsonine, a powerful inhibitor of α -mannosidases from jack beans and almonds ($K_i = 2$ and $10 \mu\text{M}$) but more specific than the natural alkaloid because it does not inhibit β -galactosidases [5]. A synthetic piperidine with the absolute configuration of β -D-galacto-hexopyranoside (5) was also submitted to the tests and this, surprisingly, showed complete inactivity against five β -galactosidases but it was a

good competitive inhibitor of β -glucosidases ($K_i = 15 \mu\text{M}$) from sweet almonds [6].

Conclusions and Perspectives

Our screening of potential glycosidase inhibitors is proceeding with many different molecules of both natural and synthetic origins, with samples from several laboratories. For the future, we are planning to enlarge the enzyme screening panel with recombinant mammalian glycosidases and glycosyltransferases.

Received: September 30, 1996

- [1] G. B. Karlsson, T.D. Butters, R.A. Dwek, F.M. Platt, *J. Biol. Chem.* **1993**, *268*, 570.
- [2] I. Pastuszak, R. Molyneux, L.F. James, A.D. Elbein, *Biochemistry* **1990**, *29*, 1886.
- [3] A. Brandi, S. Cicchi, F.M. Cordero, R. Frignoli, A. Goti, S. Picasso, P. Vogel, *J. Org. Chem.* **1995**, *60*, 6806.
- [4] Y. Chen, P. Vogel, *J. Org. Chem.* **1994**, *59*, 2487.
- [5] S. Picasso, P. Vogel, *J. Org. Chem.* **1995**, *60*, 6806.
- [6] A. Baudat, S. Picasso, P. Vogel, *Carbohydr. Res.* **1996**, *281*, 277.