doi:10.2533/chimia.2013.267

Chimia 67 (2013) 267-270 © Schweizerische Chemische Gesellschaft

[(Cp-R)M(CO)₃] (M= Re or ^{99m}Tc) Conjugates for Theranostic Receptor Targeting

Daniel Can[§], Paul Schmutz, Samer Sulieman, Bernhard Spingler, and Roger Alberto*

§SCS-DSM Award for best poster presentation

Abstract: Cyclopentadienyl complexes of ^{99m}Tc became accessible *via* a retro Diels-Alder synthetic approach of dimerized cyclopentadiene derivatives. So far, this approach was limited to derivatives comprising a carboxylic acid group, directly conjugated to the Cp-ring, leading to complexes $[(C_5H_5COCH)^{99m}Tc(CO)_3]$ and $[(C_5H_5CONH-R)^{99m}Tc(CO)_3]$, respectively. The introduction of an –NCO group *via* Curtius rearrangement and subsequent *in situ* reactions with alcohols or amines gave $[(C_5H_5NHCO-OR)_2]$ and $[(C_5H_5NHCO-NHR)_2]$. To increase the spacer lengths between the Cp-ring and the functional groups, methylene and ethylene spacers were introduced to yield $C_5H_5-CH_2COOH$ and $C_5H_5-C_2H_4COOH$ respectively. The latter Cp-derivatives reacted with $[^{99m}TcO_4)]^-$ and in the presence of CO releasing/reducing agents to the corresponding $[(C_5H_5-spacer-COOH)^{99m}Tc(CO)_3]$ complexes. The carboxylato groups can be derivatized with targeting functions, leading to structurally altered receptor binding complexes, with ^{99m}Tc for imaging and with rhenium for therapy. The nature of the ^{99m}Tc complexes was assessed by HPLC comparison with the corresponding rhenium compounds.

Keywords: Bioorganometallic chemistry \cdot Carbonic anhydrase inhibitors \cdot Radiopharmaceuticals \cdot Technetium \cdot Theranostic

Introduction

Organometallic complexes with bioactive ligands are of interest for noninvasive imaging of biological events and therapeutic treatment of diseases. While several delement cations are studied and used for therapy, ^{99m}Tc is the most prominent in nuclear medical diagnostics. It would be desirable in a theranostic sense (therapy and diagnostics), to have identical homologous compounds for combined therapy and diagnosis.^[1-3] Rhenium and technetium belong to the same traid; therefore it is possible to use respective congener compounds for matched therapy and imaging.^[4-6] While Re-based compounds can be used for therapy, the 99mTc homologs can serve as imaging agents for Single Photon Emission Computed Tomography (SPECT).^[1,2,7] $[(CP-R)M(CO)_{2}]$ -type compounds are

very stable under physiological conditions and can synthetically be treated like aromatic organic molecules. Focussing on the synthesis of ^{99m}Tc-labelled radiopharmaceuticals and the corresponding cold rhenium compounds, a variety of organic and inorganic cyclopentadienyl derivatives has been synthesized and their complexes biologically investigated.

Medicinal inorganic chemistry has progressed tremendously since the successful introduction of Cisplatin in the late 1960s. During the past 50 years, a multitude of metal-containing anticancer, anti-inflammatory, antomicrobial, antimalarial and antibacterial compounds, enzyme inhibitors and MRI-contrast agents have been exploited and investigated.^[8,9] The vast majority of clinically applied pharmaceuticals are of organic nature. Many of the organometallic or inorganic drug candidates are believed to act in the same way as Cisplatin whereas metal complexes for structural recognition of receptors are rarely found. Only recently, organometallic compounds came into the scope of such applications. The structural complexity of metal-containing compounds that are stable in biological systems was recognized to have an underestimated and unexplored potential for medicinal applications.[9-13]

Since nature has developed an enormous realm of functionally and structurally similar receptors and enzymes, selectivity for a specific target is a key criterion for the quality of an inhibitor.^[14] Meggers and coworkers showed recently that high selectivity not only depends on intermolecular interactions but also on a versatile and directed three-dimensional arrangement of different functionalities. As exemplified by organometallic protein kinase inhibitors,^[15–17] chemically inert organometallic complexes offer the opportunity to occupy biologically relevant chemical space as compared to similar inhibitors with a purely organic scaffold. Therefore, bioorganometallic complexes exhibit high potential as chemical probes.^[15,18–21]

The introduction of sterically demanding metal complexes into small molecules without affecting their biological activity is challenging. One possible strategy was pioneered by Jaouen and coworkers. In the late 1970s, Hanzlik et al. replaced a phenyl ring in phenylalanine by ferrocene and observed that this organometallic Cp⁻ sandwich-complex was accepted as a substrate in the binding pocket of phenylalanine decarboxylase.^[22] Based on this observation, Jaouen developed a variety of highly potent estrogen receptor inhibitors by replacing a phenyl ring in Tamoxifen with [(Cp-R)Re(CO)₃]. The resulting molecules retained a high binding affinity for the estrogen receptor.[12,23-28]

Combining the observations made by Meggers and by Jaouen, we studied the

^{*}Correspondence: Prof. Dr. R. Alberto University of Zürich Institute of Inorganic Chemistry Winterthurerstrasse 190 CH-8057 Zürich Tel.: +41 44 635 46 31 E-mail: ariel@aci.uzh.ch

biological activity and the selectivity of [(Cp-R)Re(CO)₃]-complexes with different targeting vectors. In order to follow the theranostic concept, we prepared the corresponding ^{99m}Tc-complexes in water. For future projects, we investigated different Cp-building blocks comprising different functional groups bound to the Cp-ring.

Results and Discussion

The derivatization of cyclopentadiene (HCp) or its complexes has a long history in organic and organometallic chemistry. Many different substituents have been described in literature.^[29–35] However, reports for coupling biologically active targeting vectors are comparably scarce. Being part of the spacer between the chelator and the biovector, the functional group attached to Cp is of great importance for a bifunctional chelator (BFC).

We reported the unexpected formation of $[(CpCOOH)^{99m}Tc(CO)_3]$ from 'Thiele's acid', the Diels-Alder dimer of HCp-COOH,^[36–38] and subsequently a general aqueous synthetic route to $[(Cp-R)^{99m}Tc(CO)_3]$ -type complexes was developed by derivatization of the acid function *via* amide coupling to biotargeting vectors. This procedure, where the carbonyl group is assumed to play the important role of a metal-anchoring group, allows a fully aqueous preparation of a variety of bioactive compounds.^[6,37,39]

Following this approach, we recently reported organometallic carbonic anhydrase inhibitors (CAI) with superior selectivity for the pharmaceutically relevant isoforms IX and XII.^[6] One of the described examples consisted of a [(CpNHCO-R) Re(CO)₃] building block derivatized with an arylsulfonamide targeting vector and showed the most pronounced selectivity profile. The corresponding ^{99m}Tc-complex however, has not yet been described, as the required compound HCp-NH₂ is still unknown.

Besides CAIs, many pharmacological active species and natural substances are aniline-based and contain the amino group in the form of amides or as internal aryl-sulfonamides. The replacement of such motives by the organometallic congener $[(CpNHCO-R)M(CO)_3]$ (M = Re or ^{99m}Tc) would therefore increase application opportunities in therapy and in diagnosis.

Coupling of a hypothetical HCp-NH₂ to carboxylic acids would yield a carbonyl group close to the HCp-ring. As shown with ketones^[36] and carboxylic acids,^[37] the carbonyl group in α -position to the HCp is an anchoring group, binding to [^{99m}Tc(OH₂)₃(CO)₃]⁺ and initializing the *retro* Diels-Alder reaction and formation of η^5 -coordinated Cp⁻-complexes. With





Scheme 1. Synthetic conditions: i) diphenylphosphoryl azide (DPPA), NEt₃, toluene 80%; ii) heated to 80 °C, toluene; iii) addition of the nucleophiles a/b, toluene.

Fig. 1. Time-resolved IR-measurement of the Curtius rearrangement of **1** to **2** (complete after 20.52 min) and the following nucleophilic addition of MeOH (after 38.51 min) to form **3a**.

Scheme 2. Synthetic conditions: i) bromoacetic acid, THF, 30-50%; ii) methyl-3-brompropionate, THF, 30-50%; iii) LiOH, THF/MeOH/ H₂O 69–74%.

Scheme 3. Synthetic conditions: i) $Na_2B_4O_7^*10H_2O$ (Borax), H_2O , 95 °C, 20%.

HCpNHCO-R, the carbonyl group would be in the β -position, still close enough to act as an anchoring group.

Aiming at the synthesis of HCp-NH₂, compounds **3a** and **3b** were synthesized by Curtius rearrangement (Scheme 1). The acid-azide **1** was obtained after treatment of Thiele's acid with diphenylphosphoryl azide in toluene. Heating to 80 °C induced the Curtius rearrangement. The resulting isocyanate reacted *in situ* with an alcohol or an amine, to form **3a** or **3b** respectively. With MeOH, the reaction was followed in a time-resolved IR-experiment, evidencing the successful transformation (Fig. 1).

Another motive in bioactive pharmaceuticals is $Ar-(CH_2)_n$ -CONHR. Organometallic analogues of this moiety could represent biomimetics for new pharmaceuticals or imaging agents.

Scheme 4. Synthetic

conditions: i) triethyl phosphonoacetate, MeOH. 80%; ii) H₂/

Pd/C, EtOAc (10a).

THF 98% (10b).

iii) LiOH, H_oO/MeOH/



Fig. 2. ORTEP representation of **8**. Relevant bond lengths (Å): Re(1)–C(1) 1.912(2), Re(1)–C(2) 1.917(2), Re(1)–C(3) 1.913(2), Re(1)–C(7) 2.299(2), Re(1)–C(6) 2.3002(19), Re(1)–C(5) 2.301(2), Re(1)–C(4) 2.3053(19), Re(1)–C(8) 2.308(2).

As aforementioned, HCp-compounds of the form C_5H_5 -COOH can be labelled as monomers or as dimers with ^{99m}Tc.^[36–38] In the ligands **5** and **7** with an extended spacer, this anchoring group is retained. The carbonyl group is now in β - or in γ -position relative to the HCp-ring, likely still close enough to serve as an anchoring group. Like Thiele's acid, **5** and **7** can be derivatized with bioactive functions.

Compounds **4** and **6** have both been described before.^[30,40] With bromoacetic acid methylester or bromopropionic acid methylester respectively, **4** and **6** were synthesized and basic hydrolysis gave **5** and **7** (Scheme 2).

The Re-complex **8** was synthesized directly in water. Similar to ^{99m}Tc-labeling of HCp-compounds, ligand **5** was reacted with $[\text{ReBr}_3(\text{CO})_3]^{2-}$ in water with Borax $(\text{Na}_2\text{B}_4\text{O}_7*10\text{H}_2\text{O})$ at 95 °C, representing one of the rare examples of an aqueous synthesis of a complex $[(\text{CpR})\text{Re}(\text{CO})_3]$ (Scheme 3). Yields were low (20%) due to a competing cluster formation reaction



Fig. 3. ORTEP representation of **10b**. Relevant bond lengths (Å) and angles (°): Re(1)–C(2) 1.910(3), Re(1)–C(3) 1.919(3), Re(1)–C(1) 1.919(3), Re(1)–C(6) 2.287(3), Re(1)–C(5) 2.291(3), Re(1)–C(7) 2.304(3), Re(1)–C(8) 2.311(3), Re(1)–C(4) 2.324(2), C(2)-Re(1)–C(1) 90.10(12), C(3)-Re(1)-C(1) 90.61(12).



of $[\text{Re}(\text{OH}_2)_3(\text{CO})_3]^+$ under basic aqueous conditions.^[41]

Complex **8** was analyzed by IR (KBr), NMR (CDCl₃), ESI-MS (CH₃OH) and elemental analysis. Crystals were grown from CH₂Cl₂/hexane and the structure was elucidated (Fig. 2).

Carboxylic acid complex 10b was not accessible along the same route by using ligand 7. Instead, 10b was obtained by Horner-Wardsworth-Emmons olefination^[42] of $[(CpCOH)Re(CO)_{3}]$.^[43] Treatment with triethyl phosphonoacetate in MeOH gave olefin 9 in 80% yield. Hydrogenation of the double bond with H₂ on Pd, followed by ester hydrolysis of **10a** with LiOH gave the desired complex 10b (Scheme 4). Crystals were grown from CH₂Cl₂/hexane and the expected structure was confirmed (Fig. 3).

Compounds **5** and **7** were subjected to labeling studies with ^{99m}Tc. Ligands were present as monomers and in mM concentrations. The reactions were studied 'all in one'. The ligand and an Isolink Kit were sealed in a vial and $[^{99m}TcO_4]^-$ was added. Alternatively, a two-step procedure was employed; $[^{99m}Tc(OH_2)_3(CO)_3]^+$ was synthesized separately and added to the ligand. Both procedures resulted in the clean formation of the ^{99m}Tc-complexes **11** and **12** respectively (Scheme 5 and 6). No relevant amounts of side products were observed in either case. The products were analyzed by coinjection with the corresponding Re



Scheme 5. Bottom-up trace: γ -trace of the one-pot labeling of **5** with [^{99m}TcO₄]⁻ and Isolink Kit. Top-down trace: UV-trace of Re-compound **8**. Quantitative and radiochemically pure conversion of [^{99m}TcO₄]⁻ to complex **11**.

complexes. Since 99mTc complexes are present in nanomol quantities only, comparison of retention times (radio- (99mTc) and UV/vis detection (Re) are the only methods to assess the nature of 99mTc compounds. Formation rates of the 99mTc products are different for ligands 5 (hydrolyzed 4) and 7 (hydrolyzed 6). With 5, formation was quantitative after 30 min (Scheme 5) whereas the reaction with 7 gave only 7% product after the same time and 62% after 5 h. Higher rates were achieved when the labeling with 7 was performed in a microwave reactor; at 130 °C and 30 min 65% product 12, 16% [99mTcO₄]and 19% [99m Tc(OH,),(CO)] + were obtained (Scheme 6).

In order to prepare organometallic CAI with ^{99m}Tc and based on **5** and **7**,^[6] compounds **13** and **14** were prepared from **8** and **10b** with rhenium. Compounds **13** and **14** extend our list of CAI, containing a methylene spacer and an ethylene spacer respectively between the carbonyl group and the Cp-ring. These methylene groups alter the distance between receptor binding motive and pendent complex and will allow a more profound insight into SARs for this important class of bioorganometallic CA targeting agents (Scheme 7).

Conclusion

Derivatives of HCp comprising a CP-NHCO-R spacer were prepared in their dimeric form from Thiele's acid by use of the Curtius rearrangement. The reaction progress was investigated by time-resolved IR-measurement. Addition of nucleophiles like alcohols or amines to isocyanate 2, generated *in situ* the compounds **3a** and **3b**.

Compounds 5 and 7 contain a carboxylic acid function in β - or in γ -position, with respect to the HCp-ring. Labelling experiments with $[^{99m}Tc(OH_2)_3(CO)_3]^+$ proved that these positions are still close enough to allow anchoring of the metal center by the carboxylic acid portion. Therefore, bringing the metal in close proximity, η^5 coordination of the HCp-portion is simplified. However, conversion rates of the ^{9m}Tc starting compounds were very different for the ligand 5 compared to 7, which is in accordance with the increasing distance of the anchoring group to the HCp-moiety. Compound 8 was synthesized in a similar fashion to the 99mTc-compound 11 using ligand 5 with $[Re(Br)_3(CO)_3]^{2-}$ in water,

2009. 4. 1930.





representing one of the rare examples of an aqueous synthesis of a $[(CpR)Re(CO)_3]$ complex. Compound **10b** was prepared using a Horner-Wardsworth-Emmons ole-fination.

Compounds 13 and 14 extend our list of CAI, containing a methylene spacer and an ethylene spacer, respectively, between the carbonyl group and the Cp-ring. These groups alter the distance between receptor binding motive and pendent complex and will allow a more profound insight into SARs for this important class of bioorganometallic CA targeting agents.

Received: March 7, 2013

- R. Alberto, in 'Bioinorganic Medicinal Chemistry', Ed.: E. Alessio, Wiley-VCH, Weinheim, 2011, pp. 253.
- [2] J. R. Dilworth, S. J. Parrott, *Chem. Soc. Rev.* **1998**, 27, 43.

- [3] D. Can, H. W. Peindy N'Dongo, B. Spingler, P. Schmutz, P. Raposinho, I. Santos, R. Alberto, *Chem. Biodiv.* 2012, 9, 1849.
- [4] V. Ozdemir, B. Williams-Jones, *Nat. Biotechnol.* 2006, 24, 1324.
- [5] F. Pene, E. Courtine, A. Cariou, J. P. Mira, *Crit. Care Med.* 2009, *37*, 850.
- [6] D. Can, B. Spingler, P. Schmutz, F. Mendes, P. Raposinho, C. Fernandes, F. Carta, A. Innocenti, I. Santos, C. T. Supuran, R. Alberto, *Angew. Chem. Int. Ed.* **2012**, *51*, 3354.
- [7] R. Alberto, J. Organomet. Chem. 2007, 692, 1179.
- [8] N. J. Farrer, P. J. Sadler, in 'Bioinorganic Medicinal Chemistry', Ed.: E. Alessio, Wiley-VCH, Weinheim, 2011, pp. 1.
- [9] G. Gasser, N. Metzler-Nolte, Curr. Opin. Chem. Biol. 2012, 16, 84.
- [10] C. Biot, G. Glorian, L. A. Maciejewski, J. S. Brocard, J. Med. Chem. 1997, 40, 3715.
- [11] L. Delhaes, C. Biot, L. Berry, P. Delcourt, L. A. Maciejewski, D. Camus, J. S. Brocard, D. Dive, *ChemBioChem* 2002, *3*, 418.
- [12] G. Jaouen, S. Top, A. Vessières, P. Pigeon, G. Leclercq, I. Laios, *Chem. Commun.* 2001, 383.

Scheme 6. Bottomup trace: γ -trace of the one-pot labeling of **7** with [^{99m}TcO₄]⁻ and Isolink Kit. Topdown trace: UV-trace of Be-compound **10b**

 Down trace: Ov-trace
 Pagano, D. S. Williams, Synlett 2007, 1177.

 of Re-compound 10b.
 [16] E. Meggers, Angew. Chem. Int. Ed. 2011, 50, 2442.

[17] E. Meggers, Curr. Opin. Chem. Biol. 2007, 11, 287.

[13] M. Patra, G. Gasser, A. Pinto, K. Merz, I. Ott, J.

[15] E. Meggers, G. E. Atilla-Gokcumen, H.

[14] S. V. Frye, Nat. Chem. Biol. 2010, 6, 159.

E. Bandow, N. Metzler-Nolte, ChemMedChem

Bregman, J. Maksimoska, S. P. Mulcahy, N.

- [18] L. Feng, Y. Geisselbrecht, S. Blanck, A. Wilbuer, G. E. Atilla-Gokcumen, P. Filippakopoulos, K. Krailing, M. A. Celik, K. Harms, J. Maksimoska, R. Marmorstein, G. Frenking, S. Knapp, L.-O. Essen, E. Meggers, J. Am. Chem. Soc. 2011, 133, 5976.
- [19] N. Metzler-Nolte, Angew. Chem. Int. Ed. 2001, 40, 1040.
- [20] U. Schatzschneider, N. Metzler-Nolte, Angew. Chem. Int. Ed. 2006, 45, 1504.
- [21] G. Gasser, I. Ott, N. Metzler-Nolte, J. Med. Chem. 2011, 54, 3.
- [22] R. P. Hanzlik, P. Soine, W. H. Soine, J. Med. Chem. 1979, 22, 424.
- [23] R. E. Mewis, S. J. Archibald, Coord. Chem. Rev. 2010, 254, 1686.
- [24] N. Metzler-Nolte, *Top. Organometal. Chem.* **2010**, *32*, 195.
- [25] K. H. Thompson, C. Orvig, *Dalton Trans.* 2006, 761.
- [26] S. J. Dougan, P. J. Sadler, Chimia 2007, 61, 704.
- [27] P. J. Dyson, G. Sava, Dalton Trans. 2006, 1929.
- [28] L. Feng, Y. Geisselbrecht, S. Blanck, A. Wilbuer, G. E. Atilla-Gokcumen, P. Filippakopoulos, K. Kraling, M. A. Celik, K. Harms, J. Maksimoska, R. Marmorstein, G. Frenking, S. Knapp, L. O. Essen, E. Meggers, J. Am. Chem. Soc. 2011, 133, 5976.
- [29] R. C. Kerber, M. J. Chick, J. Org. Chem. 1967, 32, 1329.
- [30] D. Scapens, H. Adams, T. R. Johnson, B. E. Mann, P. Sawle, R. Aqil, T. Perrior, R. Motterlini, *Dalton Trans.* 2007, 4962.
- [31] J. Boonsompat, A. Padwa, J. Org. Chem. 2011, 76, 2753.
- [32] M. H. Nantz, X. Radisson, P. L. Fuchs, Synth. Commun. 1987, 17, 55.
- [33] D. Chong, D. R. Laws, A. Nafady, P. J. Costa, A. L. Rheingold, M. J. Calhorda, W. E. Geiger, *J. Am. Chem. Soc.* 2008, *130*, 2692.
- [34] S. Top, J.-S. Lehn, P. Morel, G. Jaouen, J. Organomet. Chem. 1999, 583, 63.
- [35] M. Cais, J. Kozikowski, J. Am. Chem. Soc. 1960, 82, 5667.
- [36] J. Wald, R. Alberto, K. Ortner, L. Candreia, Angew. Chem. Int. Ed. 2001, 40, 3062.
- [37] Y. Liu, B. Spingler, P. Schmutz, R. Alberto, J. Am. Chem. Soc. 2008, 130, 1554.
- [38] T. Okuyama, Y. Ikenouchi, T. Fueno, J. Am. Chem. Soc. 1978, 100, 6162.
- [39] H. W. P. N'Dongo, Y. Liu, D. Can, P. Schmutz, B. Spingler, R. Alberto, *J. Organomet. Chem.* 2009, 694, 981.
- [40] R. L. Schaaf, C. T. Lenk, J. Org. Chem. 1964, 29, 3430.
- [41] A. Egli, K. Hegetschweiler, R. Alberto, U. Abram, R. Schibli, R. Hedinger, V. Gramlich, R. Kissner, P. A. Schubiger, *Organometallics* 1997, 16, 1833.
- [42] E. Metay, M. C. Duclos, S. Pellet-Rostaing, M. Lemaire, R. Kannappan, C. Bucher, E. Saint-Aman, C. Chaix, *Tetrahedron* 2009, 65, 672.
- [43] J. M. Heldt, N. Fischer-Durand, M. Salmain, A. Vessieres, G. Jaouen, J. Organomet. Chem. 2004, 689, 4775.